Indian J. Fish., 68(2): 76-85, 2021
DOI: 10.21077/ijf.2021.68.2.87166-10

Morphometry and molecular characterisation of semen in four carp species

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ABSTRACT

Present study aimed to compare the morphometric features of sperm and seminal protein profile of four cyprinid species *viz*. *Labeo rohita* (Hamilton, 1822), *Labeo fimbriatus* (Bloch, 1795), *Cyprinus carpio* (Linnaeus, 1758) and *Puntius carnaticus* (Jerdon, 1849) in order to elucidate quantitative characteristics of sperm morphology and seminal protein profiles within the family. Morphological studies revealed that carps have uniflagellate, acrosome less sperm with an ovoid head and irregular shaped mid piece. Size of sperm in terms of length and breadth showed significant variation among the four species studied. The sperm and seminal protein profiles of *Puntius carnaticus* differed from other three groups with the presence of proteins with molecular mass of 15 and 49 kDa and absence of four proteins equivalent to molecular mass of 13, 20, 21 and 23 kDa. Two dimensional gel electrophoresis (2D-GE) analysis of sperm protein profile between *Puntius carnaticus* and *Labe orohita* revealed presence of 237 protein spots.

Keywords: Cyprinid, Morphometry, Protein profiling, SDS-PAGE, Sperm

Introduction

Fish seed and feed are two major constraints in the growing aquaculture production and it became necessary to find out new techniques and technologies to enhance the breeding and seed production of fish for sustainable aquaculture enterprise (Jacobson *et al.,* 2005; Ayyappan *et al.,* 2011; Pandey, 2014). It has been reported in cattle that breed selection can be done based on the quality of milt and sperm proteins, which supports successful reproduction (Selvaraju *et al.,* 2009). Unlike other terrestrial animals, fish have a range of reproductive patterns (Maricchiolo *et al.,* 2004) and the sperm morphology can be varied due to the various developmental processes during evolution (Bonilla and Xu, 2008).

Structure and the molecular composition of sperm vary widely among fish and it is hard to correlate the sperm structure with taxonomy (Mattei, 1991). So far, the studies related to morphological and structural analysis of fish sperm, especially freshwater fish were done from an evolutionary point of view (Baccetti *et al.,* 1984; Mattei, 1991; Marricholo *et al.,* 2002). Recent studies shed more light on the variations of sperm even at species level. Sometimes, the general description of sperm structure and organisation of a family may not be sufficient for developing a strategy for successful breeding of fish. Quality of the milt depends on various characteristics of sperm and seminal plasma components (Migaud *et al.,* 2013). So the protein composition of seminal plasma and sperm need to be studied.

Proteomic analysis of sperm cells in animals including fish has provided information regarding the role of proteins involved in sperm physiology and functions in its various structural parts (Ciereszko *et al.,* 2012). Specific key roles of proteins on a variety of processes have been linked to reproductive performance and gamete quality. Most of the studies concentrated on fish with commercial interest aiming at improvement in broodstock management, breeding strategies and sperm cryopreservation. In addition, proteomic analysis revealed to be a good indicator of cryopreservation success by assessing specific cryo-damage. Protein profiling based on two dimensional PAGE analysis can be used to separate very closely related proteins and to get a clear picture of various proteins in cells including spermatozoa (Pandey and Mann, 2000). This helps to understand the functional role of various proteins in seminal plasma and sperm (Dietrich *et al.,* 2014). Even though artificial reproduction and cryopreservation of gametes are practiced in fish

reproduction, the proteomics analysis of milt may further improve the techniques for assuring germ cell quality. Moreover, if molecular characteristics and the functional composition of gametes and seminal plasma are known, genetical and nutritional interventions can be incorporated for enhancing the reproductive potential of brooders.

Carps such as *Labeo rohita* (LR, Indian major carp), *Labeo fimbriatus* (LF, Indian minor carp), *Cyprinus carpio* (CC, Exotic carp) and *Puntius carnaticus* (PC, Kaveri carp, barb) are representatives of Cyprinidae family, with the first two carp species widely cultured in India and common carp cultured throughout the world, while *P. caranaticus* is widely present in the peninsular region of India. The breeding season of LR is in monsoon while that of LF and PC is post-monsoon but CC can be bred throughout the year if optimum conditions are provided. Interspecific variation in quantitative sperm morphology and protein profiling of sperm and seminal plasma has provided an opportunity for developing markers for male brood traits selection and for making strategies for *ex situ* conservation of the germplasm. Additionally, a comparative study between bull and fish seminal protein profiles has contributed basic information on protein changes in sperm environment related to internal and external fertilisations. Hence, the present study aimed to generate baseline information on the structure and morphometry of sperms as well as proteins abundance in the milt and sperm during breeding season, in four cyprinid species.

Materials and methods

Sample collection

Milt was collected from 3 individuals of each species of 1 year old *L. rohita* (190-200 g), *L. fimbriatus* (180-200 g), *C. carpio* (200-230 g) and *P. carnaticus* (140-150 g) from earthen ponds of ICAR-Central Institute of Freshwater Aquaculture (ICAR-CIFA), Regional Centre, Hessarghata, Bangalore, India. The fish were injected with Ovatide (sGnRH analogues; HemaPharma, India) 0.02 ml kg^{-1} to induce breeding and the milt was collected by gentle stripping of abdomen after 6 h of injection. Utmost care was taken to avoid stress to the fish. For protein studies, milt samples were centrifuged at 11000 *g* for 15 min to separate seminal plasma from sperm cells. The sperm pellet and seminal plasma samples were snap frozen in liquid nitrogen and stored at -80°C until protein analysis. For morphometric studies 50 µl of samples were suspended in 1 ml of buffer comprising 0.484% Tris calcium chloride (TC) (39.9 mMTris, 2.6 $mMCaCl_2$) and 1.5 $mMNaN_3$. Fishes were kept in separate tank for definite period and back to experimental ponds.

Morphometry analysis

The milt was diluted (1: 200) in TC buffer and fixed by adding 2 µl of neutral buffered formalin (10%). Diluted semen samples of 10 µl were mixed with 10 µl of 3% rose bengal stain and an aliquot of 8 µl was smeared on glass slides. Smears were prepared in triplicate for all the 4 species. The slides were allowed to dry for 15-20 min and washed for one minute, dried and observed under phase contrast microscope (Nikon ECLIPSE 80 i). High resolution images of various fields were captured at 400X. In each smear, 100 sperm cells were chosen for morphometric analysis. Morphometric parameters of spermatozoa *viz*., head length, head breadth, total length, head area, head radius, head diameter, mid piece length and flagellar length/tail length, were calculated using the software NIS elements BR3.1 (Nikon ECLIPSE 80 i). The flagellar length was measured from the mid piece (dark elongated spot like structure separating the head and tail portion) end to the tip of the flagella (Fig. 1).

Extraction of protein

The sperm pellets were thawed on ice and the sperm cells were centrifuged at 8000 *g* for 5 min at 4°C and the supernatant was discarded. This washing process was repeated two times and the pellet was re-suspended in 1 ml TC buffer containing 10 µl 0.1 M phenyl methyl sulphonyl fluorides (PMSF). This step was followed by incubation at 0.1% Triton X-100 (Polyethyleneglycol 4-octyl phenyl ether) for 1 h at 4°C with intermittent vortexing at 10 min interval. After an hour, the samples were centrifuged at 13000 *g* for 30 min at 4°C and the supernatant containing sperm protein was collected in a fresh micro-centrifuge tube. The seminal plasma and sperm proteins were estimated by Bradford method using bovine serum albumin as a standard (Bradford, 1976).

Protein profiling

Protein profiling of fish samples were done using sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE) in order to identify species-specific protein variation. Based on the SDS-PAGE

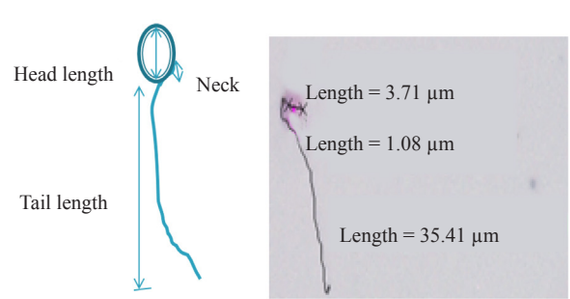

Fig. 1. Morphometry of carp sperm showing head length, tail length and neck

protein profile variation among species, distantly related 2 fish samples were subjected to the two dimensional gel electrophoresis (2DE) technique. The results were compared with that of bull sperm protein.

One-dimensional electrophoresis (SDS-PAGE)

Sperm proteins and seminal plasma protein profiles were analysed by SDS-PAGE using 4% stacking gel and 12% resolving gel (mini-PROTEAN tetra cells, Bio-Rad, Hercules, CA, USA). Sperm proteins (12 µg per lane) and seminal plasma protein (20 µg per lane) were mixed with sample buffer (0.5M trisHCl, Glycerol, 10% SDS, β-mercaptethanol, 0.05% w/v bromophenol blue) in the ratio 1:1 and denatured at 95°C for 5 min before loading. After electrophoresis, the gels were stained using silver nitrate and images were scanned in gel documentation system (SYNGENE, UK). The quantitative measurement of molecular weight, raw volume, height and quantity of the proteins were recorded from the software (Gene Tools, version 4.01.02, Syngene, UK).

Two-dimensional gel electrophoresis (*2D-GE*)

Based on morphometric analysis and SDS-PAGE profiling, two different fish species were selected for 2DE analysis. The 2D-GE was performed as per the method of O'Farrell *et al*. (1977) and D'Amours *et al*. (2010) with slight modifications using Protean IEF Cell and 2D starter kit (Bio-Rad, USA). The samples (50 µg) were mixed with rehydration buffer and ampholyte to obtain 125 µl final volume and dispensed to the isoelectric focusing (IEF) well and immobilised pH gradient IPG strips (7 cm length and a pH range of 3-10, Bio Rad) were placed on the sample for rehydration. Rehydration was performed at 50V for 16 h at 200C and IEF was performed with linear/rapid focusing conditions (250V, 500V, 1000V, 3000V, 8000V until 13000Vh). After IEF, the strips were equilibrated for half an hour in equilibration buffer (6 M Urea, 57 mM TrisHCl, 29.3% glycerol 2%) containing dithiothreitol (DTT - 2.5%) for 30 min, followed by iodoacetamide (IAA - 2%) in equilibration buffer for 30 min and rinsed in electrode buffer for 5 min at room temperature (DTT, 2.5% iodoacetamide and 2% SDS; pH 8.8). Further, the strip was loaded in 12% polyacrylamide gel, sealed with agarose (1%) with bromophenol blue and performed electrophoresis. Gels were stained using silver nitrate and scanned in gel documentation system (G BOX ichemi XR, SynGene) and analysed using Dymension software (SynGene, version 2, UK).

Statistical analysis

Morphometry data were subjected to one-way ANOVA followed by Duncan's Multiple Range Test (DMRT) using SPSS package version 16.0. The Pearson-correlation analysis was performed between 2DE gel protein profiles. The data were presented as mean±SEM and $p<0.05$ was set as level of significance..

Results

Morphometry

The average volume of milt in LR, LF, CC and PC were 3.5, 3.5, 5.5 and 1.25 ml, respectively (Table 1). CC had slight higher weight and a higher milt volume than the smaller sized PC. The morphometric features of different species are presented in Table 2 and a general structure is given in Fig 1. The total length (µm) of the sperm significantly differed ($p<0.05$) among the four species and the highest total length was observed in PC. The head length (µm) and breadth (µm) were significantly lower in LR as compared to PC and CC. The maximum head area (µm^2) was observed in CC similar to PC, but differed significantly ($p<0.05$) from LF and LR. The mid piece length (Neck) was significantly ($p<0.05$) lower in LR compared to all other groups. Head diameter (µm) was significantly higher in CC. From this study, it was observed that among the four species studied, LR and PC showed a wide difference in total length of the sperm, head length, mid-piece length, area and breadth of the head as compared to other species. The LR is closely related to LF whereas PC is closely related to CC (Table 2).

Protein analysis

Seminal plasma protein concentrations (mg ml^{-1}) in all the four species did not differ significantly ($p<0.05$) (Table 3). The plasma protein profile on SDS-PAGE exhibited 17-23 bands with molecular mass ranges equivalent to 7-212 kDa in LR, LF, CC and PC respectively (Fig. 2). The molecular weights of common bands expressed in all the four species were 67, 51, 36, 34, 26, 10 and 7. PC had additional protein bands (kDa) of 22, 54, 56, 75, 82 and 29, while LR had 43, 73, 97, 112, 131 and 180 kDa. Four protein bands equivalent to 58, 91, 123 and 212 kDa were present in CC, whereas, LF plasma proteins exhibited molecular masses of 39, 35, 61, 62, 101 and 118 kDa bands additionally (Fig. 2, Table 4).

The common carp sperm membrane protein concentration (mg ml^{-1}) was 5.01±0.89 mg ml^{-1} and differed significantly ($p<0.05$) from other species, which

Table 1. Weight of the experimental fishes and milt obtained per fish in each group

Species	Weight (g)	Milt per fish (ml)
L.rohita	190-200	3.5 (Range, 3-4)
L. fimbriatus	180-200	3.5 (Range, 3-4)
C. carpio	200-230	5.5 (Range 5-6)
P. carnaticus	140-150	1.25 (Range 1-1.5)

Table 2. Morphometric measurements (μm) of spermatozoa of four cyprinids studied

Species	HL	TL	N	Total	HB	HD	HA
LF	4.90[ab]±0.14	26.86[b]±0.99	1.27[a]±0.04	32.96[d]±1.04	4.82[ab]±0.12	4.97[b]±0.17	23.51 [b] ±2.69
LR	4.34[b]±0.14	31.12[ab]±0.75	1.05[b]±0.03	36.43[c]±0.86	4.52[b]±0.11	4.70[b]±0.12	20.56 [b]±1.36
CC	5.36[a]±0.29	33.61[a]±1.57	1.29[a]±0.11	39.36[b]±0.82	5.62[a]±0.42	5.82[a]±0.43	31.96[a]±2.29
PC	5.18[a]±0.24	35.71[a]±0.71	1.40[a]±0.06	42.30[a]±.069	5.50[a]±0.19	5.37[ab]±0.04	30.54[a]±1.44

Data expressed as Mean±SE (n=100); p<0.005. LF - *L. fimbriatus,* LR - *L. rohita,* CC - *C. carpio,* PC - *P. carnaticus.*
HL-Head length, TL-Tail length, N-Neck, HB-Breadth of sperm head, HD-Diameter of head, HA- Head area

Table 3. Protein concentration in sperm membrane and seminal plasma of the four carp species

Species	Protein concentration (mg)-SM	Protein concentration (mg ml^{-1})-SP
LR	2.08[b]±0.24	0.50±0.03
LF	2.62[b]±0.16	0.71±0.17
CC	5.01[a]±0.89	1.05±0.36
PC	2.55[b]±0.26	1.06±0.22

LR - *L. rohita*, LF - *L. fimbriatus*, CC - *C. carpio*, PC - *P. carnaticus,* SM - Sperm membrane, SP - Seminal plasma.

Table 4. Various seminal plasma proteins observed in SDS-PAGE in the four carp species

S/N	Protein (KDa)	LF	LR	PC	CC
1	6	y			
2	7	y	y	y	y
3	10	y	y	y	y
4	11	y	y		y
5	14		y	y	
6	22			y	
7	26	y	y	y	y
8	29			y	
9	34	y	y	y	
10	36	y	y	y	y
11	41		y	y	
12	47	y		y	y
13	51	y	y	y	y
14	59	y	y		
15	39	y			
16	35	y			
17	43		y		
18	67	y	y	y	y
19	61	y		y	y
21	224	y			
22	118	y			
23	101	y			
24	54			y	
25	56			y	
26	75			y	
27	279		y		
28	264		y		
29	180		y		
30	131		y		
31	122		y		
32	112		y		
33	97		y		
34	58				y
35	91				y
36	123				y
37	212				y
38	82			y	

LR - *L. rohita*, LF - *L. fimbriatus*, CC - *C. carpio*, PC - *P. carnaticus,* y - yes, shows the presence of proteins.

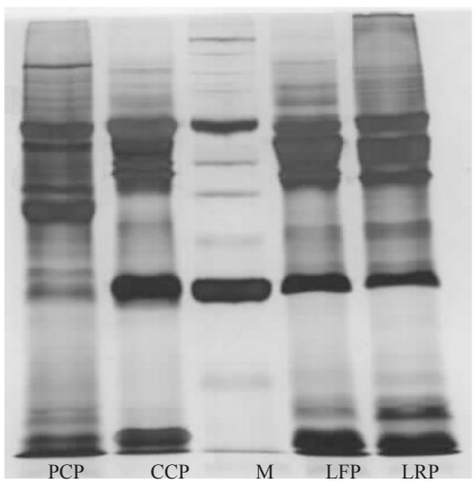

Fig. 2. Molecular weight (kDa) of proteins in seminal plasma of four cyprinid species revealed by SDS-PAGE. Gels were stained with silver staining. LRP - *L. rohita*, LFP - *L. fimbriatus*, CCP-common carp, PCP - *P. carnaticus.* Numbers indicate the molecular weight of the marker (middle) in kDa

were 2.08±0.24, 2.62±0.16 and 2.55±0.26 in LR, LF and PC, respectively. However, there was no significant difference in seminal plasma protein concentration in all the four cyprinids. Analysis of sperm protein through SDS-PAGE showed that there was a difference in abundance of the proteins in these carps. The proteins equivalent to molecular weight (kDa) of 64, 61, 58, 56, 45, 41, 36, 32, 27 and 18 expressed ubiquitously in four species. However, 66 kDa and 7 KDa proteins were expressed in all species except LF. Similarly, 52 kDa and 29 kDa were not expressed in CC, while 20 kDa, 21 kDa, 23 kDa and 13 kDa were not found in PC (Fig. 3, Table 5).

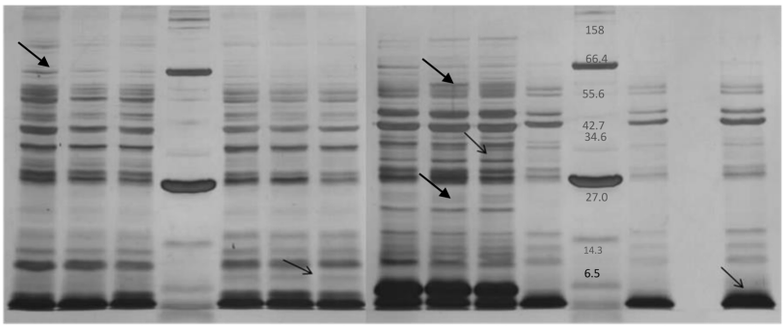

Fig. 3. Molecular weight (kDa) of proteins in sperm membrane protein of four cyprinid species revealed by SDS-PAGE. Gels were stained with silver staining. LRP-*L. rohita*, LFP-*L. fimbriatus*, CCP-common carp, PCP- *P. carnaticus.* Numbers indicate the molecular weight of the marker in kDa

Table 5. Various sperm membrane proteins observed in the four carp species

S/N	Protein (KDa)	LF	LR	PC	CC
1	237		y		
2	129			y	y
3	124		y		
4	115	y	y	y	
5	109	y			
6	105		y	y	
7	95		y	y	
8	81	y			
9	66		y	y	y
10	64	y	y	y	y
11	61	y	y	y	y
12	58	y	y		
13	58	y	y	y	y
14	56	Y	y	y	y
15	52	y	y	y	
16	49			y	
17	47	y	y		
18	45	y	y	y	y
19	41	y	y	y	y
20	37				y
21	36	y	y	y	y
22	33	y			
23	32	y	y	y	y
24	29	y	y	y	
25	27	y	y	y	y
26	25				y
27	24				y
28	23	y	y		y
29	21	y	y		Y
30	20	y	y		y
31	18	y	y	y	y
32	17				y
33	15			y	
34	13	y	y		y
35	9	y	y		
36	7		y	y	y
37	4	y		y	

LR - *L. rohita*, LF - *L. fimbriatus*, CC - *C. carpio*, PC - *P. carnaticus,* y - yes; shows the presence of proteins

The percentage abundance of individual proteins were observed as 25 and 23% for molecular mass of 32 and 41 kDa, respectively, in LF, whereas the same for 45 kDa was 22% in LR, 24% for 45 kDa in PC and 56 and 64 kDa accounted for 15 and 17%, respectively, in CC (Fig. 5).

Proteins in bull and fish seminal plasma

Bull seminal plasma showed 18 protein bands with molecular masses ranging from 8-68 kDa, while in fish it was 23 bands ranging from 7-212 kDa (Fig. 4). The maximum abundance of 46 kDa protein was observed in bull, whereas in fish such abundance was recorded for protein band equivalent to 26 kDa. The maximum number of sperm protein was 18 and 27 in bull and fish samples respectively (Fig. 4).

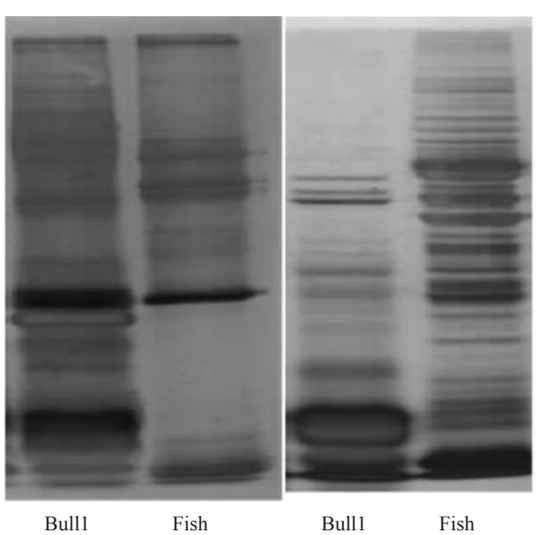

Fig. 4. Molecular weight (kDa) of proteins in bull sperm membrane and seminal plasma revealed by SDS-PAGE. Gels were stained by silver staining

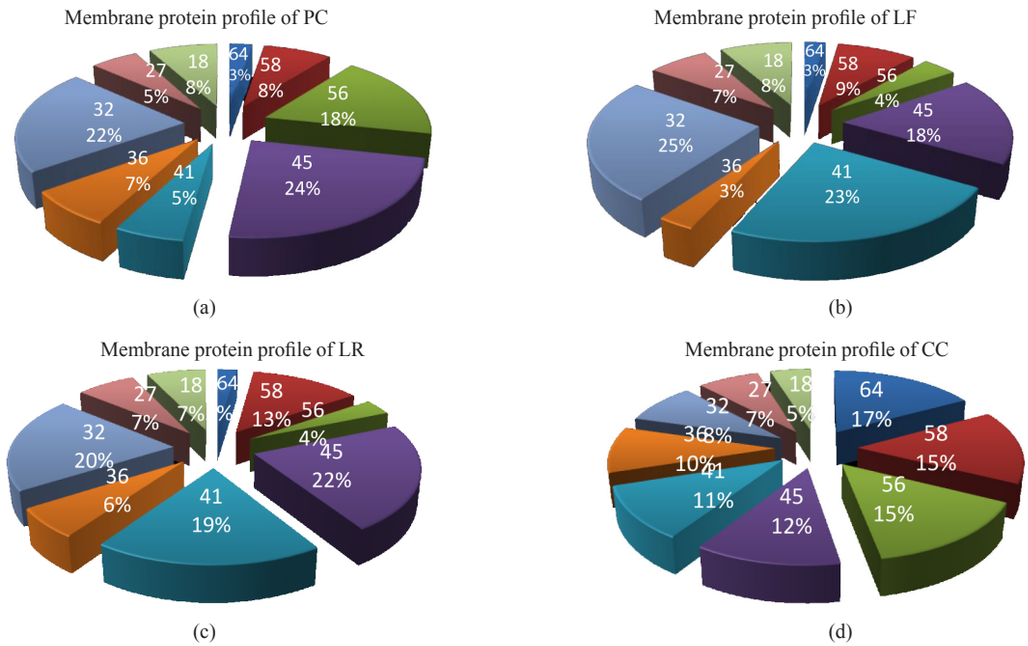

Fig. 5. Graphs showing the quantity of expressed sperm protein and membrane protein profile of (a) PC, (b) LF, (c) LR and (d) CC

Two-dimensional gel electrophoresis (2D-GE)

As morphometric and SOS-PAGE analysis showed marked quantitative and qualitative variation among the species, two-dimensional analyses were performed for PC and LR. On 2D-GE study, we observed 237 common spots in two species, such as, spot no.193 (Fig. 6). Even though 196 spots were expressed almost equally in both species, 71 of them were up-regulated in PC compared to LR (Fig. 6). The correlation co-efficient and co-efficient of determination values were calculated as 0.84 and 73% respectively (Fig. 7). Some lower molecular weight proteins with varied pI such as, 10.57 kDa, pI 9.91 and 8.60 kDa, pI 9.30 were present only in LR, while PC showed similar smaller proteins of molecular masses of 23.62 kDa, pI 4.64 and 22.7 kDa, pI 6.38.

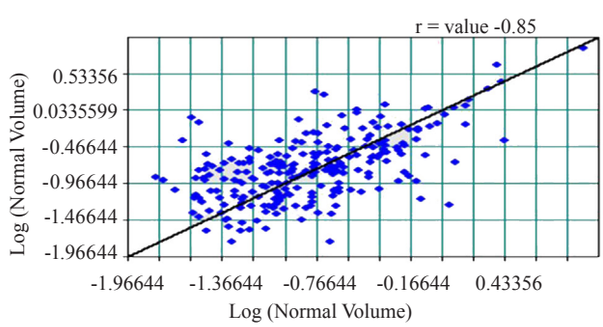

Fig. 7. Correlation chart of spots observed in 2D-GE analysis of *L. rohita* (Left) and *P. carnaticus* (right) sperm membrane protein

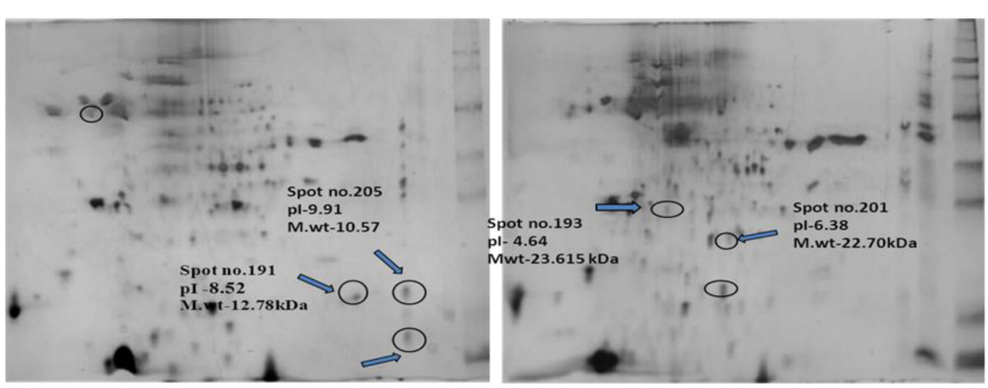

Fig. 6. 2D-GE analysis of *L. rohita* (Left) and *P. carnaticus* (right) sperm membrane protein. Marked spots correspond to examples of proteins identified from 2D-GE which are differentially expressed in both the species

Discussion

The studies related to intervention in reproduction require detailed understanding of the morphology of gametes. Studies on morphology of fish sperm especially of freshwater fish are scanty. Unlike terrestrial animals, fish show variety of reproductive patterns and most of them have external fertilisation (Tabosky, 1998). So there is ample differences in the structure and morphometry of sperm in fish when compared to cattle and human sperm. This might be the reason for higher expression of higher molecular weight proteins in both seminal plasma and sperm membrane proteins of fish compared to bull seminal samples in our study. In the present study we observed that the semen yield from different carp species were different. Total volume of milt from *C. carpio* after induction with Ovatide was greater than 6 ml while that of *L. rohita* and *L. fimbriatus* was 3.5 ml. Khan *et al.* (1992) reported similar semen yield (3.63 ml kg^{-1} body weight) from *L. rohita* injected with pituitary gland extract. Similarly, milt yield from *P. carnaticus* without hormonal induction was 1.6 ml.

Most studies on sperm cytoskeleton are focused on the flagellar axoneme. Morphometric analysis of fish sperm revealed that unlike terrestrial animals, fishes have an ovoid head and flagella connected by a mid-piece. This cell lacks acrosome and possesses a flagellum attached to one side of the head region through the neck giving a "9" shaped structure to the sperm. These characteristics were similar to those reported in carps by Mattei (1970) and Stanley (1971). In most fish species, the flagellum axoneme bears the typical arrangement of nine pairs of peripheral microtubules and one pair of central microtubules, although some species such as eel (*Anguilla anguilla*) do not possess the central microtubules (Gibbons *et al.,* 1983). This structure helps the spermatozoa travel through a narrow micropyle to reach the egg plasma membrane (Morisawa, 1995).

The plasma membrane of the spermatozoa head tightly overlays the nucleus and only a thin cytoplasmic layer remain between the plasma membrane and the nucleus. The shape of the sperm head varies with shape of the nucleus. Most nuclei show an invagination in which the axoneme will be anchored. As a consequence, nucleus shape will determine the strength of flagellar attachment to the head (Bobe and Labbe, 2010). It is reported that relatively small sperm head is observed in fishes (Johnson *et al.,* 1983). The present study showed that sperm head length varies between 4.3 and 5.36 µm with breadth ranging from 4.52 to 5.62 µm in all the four species unlike the Atlantic eel, sturgeon and paddle fish sperm, which have elongated sperm head (Shaliutina, 2014). Different sperm shapes were observed in fishes, for example, laterally flattened (*Perca fluviatilis*), banana shape (Atlantic eel) and elongated (*Mimagoniates barberi)* and spherical in Mediterranean rainbow wrasse (Lahnsteiner and Patzner, 1997; Islam and Akhter, 2011). The flagellar length in fish varies from 2.6 µm in coho salmon to 94 µm in Channel catfish (Jaspers *et al.,* 1976; Islam and Akhter, 2011). Our study showed flagellar length of four cyprinids between 26.86 and 35.71 µm (Table 1), which was relatively within the range of 36 to 60 µm reported in same family (Baccetti, 1984; Islam and Akhter, 2011). The coiling in flagellar tip occurs on dilution of carp spermatozoa in freshwater (Dreanno *et al.,* 1999a). The reduction in flagellar length affects the motility of the sperm and it is determined by the ATP exhaustion of the mid piece mitochondria (Cosson, 2008).

Comparison between bull and fish seminal plasma and sperm membrane protein

Seminal plasma proteins of animals can be originated from blood as well as seminal plasma specific proteins. Comparison between fish and bull seminal plasma proteins revealed that the higher molecular weight proteins were more abundant in fishes compared to bull seminal plasma, which was dominated with lower molecular weight proteins. Similarly, abundance of high molecular weight proteins was significantly higher in fish sperm membrane. This revealed the importance of higher molecular weight proteins (above 60kDa) for external fertilisation.

Seminal plasma and sperm membrane protein profiles among fishes

Proteins in seminal plasma are involved in the protection of spermatozoa and maintaining spermatozoa in the quiescent state, providing adequate levels of sperm nutrients for various physiological processes and protecting sperm against damage caused by proteolytic or oxidative attacks (Ciereszko *et al.,* 2000). Study on seminal plasma is very critical to optimise the protocol for artificial fertilisation, storage and handling of sperm (Alavi *et al.,* 2008). Moreover, a very few studies reported the relationship between fertilisation and seminal plasma concentration (Aas *et al.,* 1991). Some of those proteins were identified as being involved in cell metabolism, oxido-reductase activity and signal transduction, membrane trafficking, organisation and cell movement (Li *et al.,* 2010). Among species, there were distinct variations with the prominence of some species-specific bands in SDS-PAGE. *P. carnaticus* exhibited distinctly different pattern in sperm membrane and seminal plasma protein profile than other three species.

The seminal plasma proteins analysis revealed omnipresence of 7 bands among species with highest abundance of 7 kDa proteins except in common carp, in

which 26 kDa was highly expressed. In another study, the 12 proteins identified in rainbow trout (16-135 kDa) and 65 and 54 kDa were dominant among them (Lahnsteiner, 2007). In most of the cyprinid species, 16kDa band was present in seminal plasma and may be responsible for energy supply and protection of sperm as reported in rainbow trout (Lahnsteiner, 2007). Asadpour *et al.* (2013) elucidated only nine proteins with molecular weights ranging from 11.74 to 68.12kDa in rainbow trout. Such variation in molecular masses may be attributed to the major defense against reactive oxygen species (ROS), owing to the low content of cytoplasm in spermatozoa (Shiva *et al.,* 2011). The present study reveals quantitative and qualitative variation in sperm membrane proteins; LR showed presence of 28 bands of proteins ranging from 7-243 kDa, while 23 bands of proteins in a range of 4-247 kDa were present in LF. PC and CC exhibited 16 and 30 bands in the range of 4-67 kDa and 6-104 kDa, respectively. The observed variation in number and abundance of sperm membrane proteins may be either species specific (Kowalski *et al.,* 2003) or due to various environmental, nutritional, seasonal and sampling conditions and variation in broodstock strains (Yue *et al.,* 2009). In addition, variation in mode of reproduction among the species may contribute to such changes in sperm plasma membrane proteins responsible for the initiation of flagellar beating upon sperm release in water (Tubbs and Thomas, 2008), gamete fusion (Yu *et al.,* 2002) and fertilisation (Beck *et al.,* 1992).

Two-dimensional analysis (2D-GE) of LR and PC

In the present study, both morphometric and SDS-PAGE revealed a significant difference among various species and LR and PC were mostly unrelated among the four carps. So we conducted further study to differentiate closely related proteins based on two-dimensional comparison of sperm membrane protein as we could observe wide variation among species in the seminal plasma protein in our SDS PAGE study. Moreover, there are several studies on seminal plasma proteins, which reported that there is a variation in seminal plasma protein in various stages of reproduction within species (Li *et al.,* 2009, Shaliutina, 2013). The seminal plasma proteins/ molecules are mainly involved in protection and mobility of sperm in fishes when it is inside the body (Cosson *et al.,* 1997; Borges *et al.,* 2005), but outside the body, osmolality of environment plays major role in activation and mobility of sperm. A first time comparative analysis of blood proteome and seminal proteome using 2D electrophoresis reported 137 proteins in seminal proteome in which most of them were newly identified (Deitrich *et al.,* 2014). In the present study, we detected that 237 spots were common between the two species. Even though 196 spots were expressed equally in both species, 71 of them were up or down regulated in either of the species. For improving the gamete quality during storage, further identification of these proteins need to be carried out to gain insight into their biological functions like signal transduction, ion binding, embryonic developmental processes, transportation of micro and macro molecules.

In conclusion, morphological studies indicated that the sperm of carps are acrosome less, uniflagellate with an ovoid head and irregular shaped mid piece. However, the species belonging to same family showed significant sperm size variation. The sperm and seminal protein profiles of *P. carnaticus* differed from other three groups with the presence and absence of certain proteins. The 2D-GE analysis of sperm protein profile between *P. carnaticus* and *L. rohita* revealed presence of 237 protein spots. Therefore, this morphometric feature of the species gives a baseline data for further study. In addition, the study observed a higher expression of higher molecular weight proteins in both seminal plasma and sperm membrane proteins of fish compared to bull seminal samples. Seminal plasma proteins and sperm proteins have to be further studied to understand the plasma specific protein and those proteins which can be specifically helping the reproductive viability of sperm.

Acknowledgements

We sincerely acknowledge the Directors of ICAR-NIANP, Bangalore, ICAR-CIFE, Mumbai and ICAR-CIFA, Bhubaneswar for their support in conducting this research work.

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Date of Receipt : 20.02.2019
Date of Acceptance : 18.04.2021